

Singapore designs tool to investigate hospital borne bacterial infections

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Scalable CRISPRi system allows scientists to identify and tackle causes of *E. faecalis*-related diseases and drug resistance

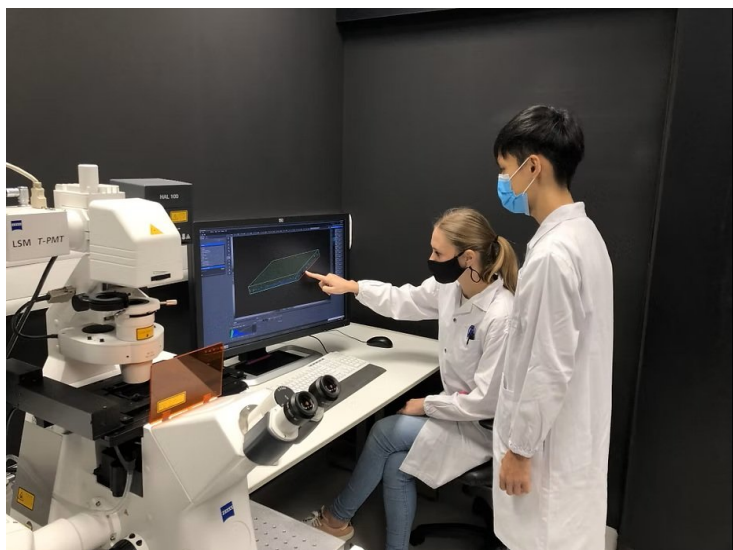


Image Caption: SMART Postdoc Dr Irina Afonina and NTU PhD Student Jerome Chua use CRISPRi technology to understand biofilm formation in *Enterococcus faecalis*

Researchers from the [Antimicrobial Resistance](#) (AMR) Interdisciplinary Research Group (IRG) at [Singapore-MIT Alliance for Research and Technology](#) (SMART), MIT's research enterprise in Singapore, and Nanyang Technological University (NTU) have developed a tool using CRISPRi technology that can help understand and prevent biofilm development, drug resistance, and other physiological behaviours of bacteria such as *Enterococcus faecalis*.

E. faecalis, a bacteria found in the human gut, is one of the most prevalent causes of hospital-associated infections and can lead to a variety of multidrug-resistant, life-threatening infections including bacteraemia (bloodstream infection), endocarditis (infection of the heart), catheter-associated urinary tract infection and wound infections.

However, current methods for understanding and preventing *E. faecalis* biofilm formation and development are labour-intensive and time-consuming. The SMART AMR research team designed an easily modifiable genetic technique that allows rapid and efficient silencing of bacteria genes to prevent infections.

In a paper titled "[Multiplex CRISPRi System Enables the Study of Stage-Specific Biofilm Genetic Requirements in *Enterococcus faecalis*](#)" published in the journal *mBio*, the researchers explain the scalable dual-vector nisin-inducible CRISPRi system which can identify genes that allow bacteria like *E. faecalis* to form biofilms, cause infections, acquire antibiotic resistance, and evade the host immune system. The team combined CRISPRi technology with rapid DNA assembly under controllable promoters, which enables rapid silencing of single or multiple genes, to investigate nearly any aspect of enterococcal biology.

"Infections caused by *E. faecalis* are usually antibiotic tolerant and more difficult to treat, rendering them a significant public health threat," says Dr Irina Afonina, Postdoctoral Associate at SMART AMR and lead author of the paper. "Identifying the genes that are involved in these bacterial processes can help us discover new drug targets or propose antimicrobial strategies to effectively treat such infections and overcome antimicrobial resistance."

The tool enables rapid and efficient investigation of a wide range of aspects of enterococcal biology and pathogenesis, host-bacterium interactions, and interspecies communication. The method can be scaled up to simultaneously silence multiple bacterial genes or perform full-genome studies.

"The system we designed enables us to easily interrogate various stages during the bacterial biofilm developmental cycle of *E. faecalis*. By selectively silencing certain genes in pre-formed, mature biofilms, we can erode the biofilm and force it to disperse." says SMART AMR Principal Investigator and NTU Associate Professor Kimberly Kline

The scalable CRISPRi system uses high-throughput screens which can allow for rapid identification of gene combinations to be simultaneously targeted for novel and efficient antimicrobial combinatorial therapies.

The idea behind SMART's inducible CRISPRi system was conceived by Professor Kline and SMART AMR Principal Investigator Professor Timothy Lu, while Dr Afonina developed and delivered the genetic tool.

The research is carried out by SMART and supported by the National Research Foundation (NRF) Singapore under its Campus for Research Excellence And Technological Enterprise (CREATE) programme.